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Curcumin and kidney protection; current findings and new concepts

Hamid Nasri¹, Zahra Abedi-Gheshlaghi², Mahmoud Rafieian-Kopaei^{3*}

¹Department of Internal Medicine, Isfahan University of Medical Sciences, Isfahan, Iran

²Nickan Research Institute, Isfahan, Iran

³Medical Plants Research Center, Shahrekord University of Medical Sciences, Shahrekord, Iran

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ABSTRACT

According to various epidemiological data, renal failure, either acute or chronic is a serious health and economical problem throughout the world. Curcumin as a member of ginger family (Zingiberaceae), is one of three curcuminoids present in turmeric and the necessity to develop kidney protective modalities sets the eyes in compounds such as curcumin, which has been used in the traditional medicine, specifically because its protective effects against kidney damage. It has been recognized that curcumin is a bifunctional antioxidant; it applies antioxidant activity in a direct and an indirect ways through scavenging reactive oxygen species and provoking an antioxidant response. Curcumin by its bifunctional antioxidant has the capability to react directly with reactive species and to stimulate up-regulation of several antioxidant and cytoprotective proteins. Thus, according to previous studies curcumin due to its properties, was effective to ameliorate the kidney damages and in this review, we will try to explain these abilities of curcumin.

Core tip: Curcumin is a free radical scavenger due to its antioxidant capacity and many experimental investigations reported its ameliorative effect in acute and chronic kidney disease such as diabetic kidney disease, nephrotoxicity and renal ischemia.

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Introduction

Chronic renal failure is a complication defined as progressive disturbance of kidney function over time and is characterized by abolishing the ability of the body to remove soluble waste resulting in the accumulation of "uremic toxins" (1-3). While, acute kidney injury is commonly described as a sudden decline in kidney function, its clinical manifestation appears as a reversible acute rise in nitrogen waste products, measured as blood urea nitrogen and serum creatinine levels, over a period of hours to weeks (1-3). Now, it is well detected that chronic renal failure is an inflammatory illness and uremic toxins have the main role in creating the inflammatory milieu (1,2). Kidney fibrosis is generally considered as activation and proliferation of interstitial area and by excessive synthesis and aggregation of extracellular matrix components, containing fibronectin and collagen. Likewise, the phenotypic changes that attribute to acute renal failure comprise inflammatory response and oxidative stress. Slowing the rate of progression of chronic renal failure and promoting the recovery of acute kidney injury is a critical part of the management of these two disease (2-4). Oxidative stress describes an imbalance between formation of reactive oxygen species

and antioxidative defense mechanisms (1-5). Uremia is associated with increased oxidative stress, and treatment of uremic individuals by hemodialysis or peritoneal dialysis has been proposed to particularly attribute to oxidative stress and diminished antioxidant levels in these groups. Loss or insufficiency of antioxidant activity might also attribute to augmented oxidative stress in uremia (1-8).

Materials and Methods

This review article discusses kidney related aspects of curcumin including its protective properties in acute and chronic renal disease and diabetic nephropathy. We report the available evidences of clinical outcomes of clinical or experimental investigations with this drug. For this review, we used a variety of sources by searching through PubMed, EMBASE, Scopus and directory of open access journals (DOAJ). The search was conducted using combinations of the following key words and or their equivalents; acute kidney injury, curcumin, antioxidant, nephrotoxicity and chronic renal failure.

Oxidative stress generation in kidney

Curcumin is a phenolic substance with chemical formula C₂₁H₂₀O₆ which is extracted from *Curcuma longa*

*Corresponding author: Prof. Mahmoud Rafieian-Kopaei,
Email: rafieian@skums.ac.ir

rhizomes is frequently used in various parts of the world (1,7-10). Furthermore, it is one of curcuminoids present in turmeric and is responsible for the yellow color of turmeric. Curcuminoids are a rich source of phenolic compounds and curcumin as the major component of turmeric has been shown antioxidant, anti-tumor, antimicrobial, anti-inflammatory and antimutagenic activities. Curcumin is soluble in acetone, ethanol and dimethylsulfoxide but insoluble in water and ether. In addition, it is rapidly conjugated and metabolized and its chemical structure for the first time, was identified in 1910 by Lampe and Milobedzka (1,7-10). Several studies have shown the ability of curcumin to prevent lipid peroxidation as a significant key in process of many diseases. In addition, curcumin is a free radical scavenger due to its antioxidant capacity and many experimental investigations reported the ameliorative effect of curcumin in acute and chronic kidney problems (1,5-12). Therefore, the aim of this study is to explain the protection effects of curcumin against kidney problems. It is well recognized that reactive oxygen species like hypochlorous acid (HOCl), or hydrogen peroxide (H_2O_2) and free radicals like superoxide (O_2^-), hydroxyl radical (OH $\cdot$), and nitric oxide (NO $\cdot$), are constantly formed in vivo (1-14). Hence, the findings of reactive oxygen species by itself does not yet outline oxidative stress, however, in a condition where antioxidative protection mechanisms are weakened, cause the imbalance between formation of reactive oxygen species and protection mechanisms which generates oxidative stress (12-16). Kidney sources for reactive oxygen species are vascular cells, activated macrophages, various glomerular cells. The balance in formation of reactive oxygen species and antioxidative protective mechanisms depends on the activity of enzymes like catalase, NO-synthase, superoxide dismutases and glutathione peroxidase. This stability, by the way, is rather fragile, hard to predict, and intensely dependent on environmental situation. Various cellular enzymes, comprising mitochondrial lipoxygenase, oxidases, cyclooxygenase, myeloperoxidase, xanthine oxidase, NADPH oxidase and in the case of L-arginine or tetrahydrobiopterin depletion, NO-synthase have been recognized as cellular sources of reactive oxygen species formation (1-16). Reactive oxygen species are also considered to contribute to the pathogenesis of ischemia-reperfusion damage (1-4,15-18). Likewise, oxidative stress mediates an extensive range of kidney impairment, from acute kidney disease, hyperlipidemia, rhabdomyolysis, obstructive nephropathy and glomerular injury which lead to chronic kidney disease and hemodialysis (12-18). Endogenous complexes in cells can be classified as enzymatic antioxidants and non-enzymatic antioxidants (12-18). The non-enzymatic antioxidants are also separated to nutrient antioxidants and metabolic antioxidants. Nutrient antioxidants, which belong to exogenous antioxidants, are compounds that cannot be produced in the body and should be provided by foods or supplements (10-19).

Moreover, due to their impact on cell cycle regulation, oxygen radicals may attribute to hypertrophy of tubular cells (38-40). Moreover, oxidative stress has role as a significant cofactor attributing to endothelial dysfunction, atherosclerosis, inflammation and glomerulosclerosis (16-20). When, reactive oxygen species are released by the mitochondrial respiratory chain can damage biomolecules like proteins, lipids and nucleic acids (11-19). To prevent the injury, antioxidant protection has evolved to remove most of these oxidant agents (1-7). Even when a balance between oxidative injury and protective mechanisms is usually reserved, there are some specific conditions in which the excessive production of free radicals, or deficiencies in antioxidant protection, results in the appearance of oxidative stress (15-21). While renal diseases patients live under predominantly various pro-oxidative situations, thus, they might be particular profit of antioxidant therapy (12-20). For many years, there have been investigations based on the application of natural compounds herbal-derived as potential therapeutic substances for various illnesses in humans (10-22).

Kidney protective effect of curcumin

It has been recognized that curcumin is a bifunctional antioxidant; it applies antioxidant activity in a direct and an indirect way through scavenging reactive oxygen species and provoking an antioxidant response. In summary, curcumin by its bifunctional antioxidant has the capability to react directly with reactive species and to stimulate an up-regulation of several antioxidant and cytoprotective proteins. Curcumin is able to scavenge superoxide anion (O_2^-), H_2O_2 , singlet oxygen, peroxyxynitrite, hydroxyl radicals (OH $\cdot$), nitric oxide and peroxy radicals (ROO $\cdot$) (2-28). Additionally, aspects such as the presence of phenolic groups in the structure of curcumin describe its capability to react with reactive nitrogen species and reactive oxygen species and it may probably be one of the mechanisms by which curcumin treatment keeps the epithelial cells of kidney tubules from oxidative damage induced by H_2O_2 . Moreover, indirect antioxidant ability of curcumin is described by its capability to stimulate the expression of cytoprotective proteins like superoxide dismutase, glutathione reductase, catalase (14-26), glutathione peroxidase (1,16-25), glutathione-S-transferase, and γ -glutamylcysteine ligase (14-24). Likewise, it has been detected that curcumin can enhance the concentration and synthesis of reduced glutathione in various cells containing astrocytes and neurons by induction of γ -glutamylcysteine ligase. In fact, recent investigations have shown that antioxidant activity may be exerted in various in vitro and in vivo models such as preventing lipid peroxidation in a variety of cells like erythrocytes, brain, liver, liposomes and macrophages, where peroxidation is brought by Fenton's reagent, in addition to hydrogen peroxide (H_2O_2), 2,2'-azo-bis(2-amidino-propane) hydrochloride (AAPH) and metals (15-29). Kidney fibrosis is largely identified by activation and proliferation of interstitial renal fibroblasts and by

extreme synthesis and accumulation of extracellular matrix substances, comprising fibronectin and collagen (25-38). Recent studies have revealed that treatment by curcumin decreases the collection of type I collagen and fibronectin in the kidneys of animals with unilateral ureteral obstruction. Activation of rat kidney interstitial fibroblasts was induced by transforming growth factor β . Curcumin treatment inhibited fibroblast proliferation and the cell cycle was arrested in the G1 phase. Curcumin treatment upregulated the expression of peroxisome proliferator-activated receptor gamma (30-38).

Diabetic kidney disease is one of the major causes of end-stage kidney failure (35-40). Diabetic kidney disease is defined by the presence of hyperfiltration, glomerular hypertrophy, mesangial matrix expansion, and increased expression of extracellular matrix proteins, tubular albuminuria (31-42). It involves several profibrotic factors like connective tissue growth factor and transforming growth factor β (1,38-45). The influence of curcumin on diabetic kidney disease has been detected by attenuation of proteinuria and improvement of creatinine clearance in rats. Accordingly, curcumin decreases oxidative stress by reducing levels of subunits of nicotinamide adenine dinucleotide phosphate oxidase that catalyzes the synthesis of O_2^- . Furthermore, it has increased the activity of the antioxidant enzyme glutathione peroxidase (31-42). More recent investigations have shown that the kidney protective property of curcumin was related to the down-regulation of the profibrotic cytokines vascular endothelial growth factor, transforming growth factor β , and osteopontin and extracellular matrix proteins fibronectin and collagen IV. Besides, it was detected a reduction in the development of structural injury detected by lower glomerulosclerosis index, tubules and interstitial fibrosis and arteriolopathy. These properties, may be in part, conducted by inhibition of protein kinase C- β (40-45). On the other hand, the protective influence of curcumin in diabetic kidney disease has also been related to the prevention of renal triglyceride buildup (40-48). More recent studies on administration of curcumin in diabetic kidney disease showed reduction of inflammatory kidney response by the attenuation of proinflammatory cytokines like tumor necrosis factor- α and monocyte chemoattractant protein-1. Additionally by attenuation of kidney macrophage infiltration, expression of the profibrotic cytokine TGF- β , inducible nitric oxide synthase (iNOS), and cyclooxygenase-2 (COX-2) could ameliorate the diabetic kidney disease (1-6,30-39).

Acute renal failure is a public health problem with an annual mortality rate which exceeds those of breast and prostate carcinoma, cardiac failure, and diabetes (42-50). After a toxic injury or an ischemic episode, dysfunction and the loss of renal tubular epithelial cells play a central role in the evolution of acute renal failure. In response to an injurious insult, tubular cells fail cytoskeletal integrity and cellular polarity, due to the dislocation of membrane proteins resulting in the disruption of cell-matrix and cell-cell integration (31-45). Central to tubular

damage is mitochondrial dysregulation, established in a reduction in cell respiration and ATP production. Mitochondrial activities rest on a complex molecular machinery delicately tuned and balanced through regulatory proteins of the two opposing processes fission and fusion (48-52). The influence of curcumin on acute renal failure has been detected, too. In this condition, curcumin was able to significantly attenuate the reduction of serum glutathione peroxidase and the increase of the malondialdehyde concentration, nitric oxide and protein carbonyl content in kidney (50-53). Importantly, the kidney protective property of curcumin is associated to maintenance of function and redox balance of mitochondria. These findings contribute to the protecting property of curcumin in the kidney to the diminution of the master regulator of antioxidant response nuclear factor erythroid-derived 2, attenuation of inflammatory response, inhibition of mitochondrial dysfunction, preservation of antioxidant enzymes and inhibition of oxidative stress (25-31). One study by Bayrak et al, showed the protective effect of curcumin against ischemia/reperfusion injury in rat kidneys. In this study, treatment with 200 mg/kg of curcumin daily for 7 days, significantly attenuated the levels of nitric oxide, protein carbonyl and malondialdehyde content in kidneys and urea, glutathione peroxidase and cystatin C levels in serum (12-18, 50-64).

Nephrotoxicity induced by chemicals

Kidney is a common target for toxicity, due to some chemical. A number of researchers have reported that curcumin can attenuate the renal injuries induced by some chemicals such as heavy metals. Heavy metal is a term that refers to any metallic chemical element and has a relatively high density and is toxic or poisonous at low concentrations. Several studies indicated the ameliorated effects of curcumin on heavy metals-induced nephrotoxicity such as cadmium, mercury and chromium. Cadmium can increase lipid peroxidation and induces oxidative stress in a biological system may be due to alterations in the antioxidant defense system (58-65). One experimental study showed the protective role of curcumin on cadmium-induced nephrotoxicity in rats. In this study male Wistar rats were treated once daily by oral gavage for 5 days and divided into 4 groups including control rats, cadmium acetate (200 mg/kg), curcumin (250 mg/kg) and in group four, rats pretreated with 250 mg/kg of curcumin for 1 hour before administration with cadmium acetate. Cadmium caused an increase in level of renal lipid peroxidation and reduced glutathione. Also the hydropic swelling and hypertrophy of proximal tubular cells were observed in renal cortex of rats which treated with cadmium. The results revealed that pretreatment with curcumin reduced both biochemical and histological alterations induced by cadmium (66). Mercury as a heavy silvery-white metal can induce nephrotoxicity via an increase in oxidative stress parameters including glutathione and lipid peroxidation levels and change in

level of catalase, superoxide dismutase and glutathione peroxidase activities in kidney. Treatment with curcumin (80 mg/kg/d) for 3 days was effective for protection against mercury-induced oxidative stress parameters and serum biochemical changes in the kidney of rats. Moreover, the concentration of mercury in the tissues was decreased after the pre/post-treatment with curcumin (66).

Potassium dichromate ($K_2Cr_2O_7$) is an inorganic chemical reagent, most commonly used as an oxidizing agent. As with all hexavalent chromium compounds, it is harmful to health and can induce nephrotoxicity in animals and humans. Molina-Jijón et al (67) showed that use of curcumin was effective in protecting of $K_2Cr_2O_7$ -induced renal oxidant damage by a mitochondrial pathway. In this study male Wistar rats, were divided into 10 groups. The treatment with curcumin was including 3 different schemes, 1) complete treatment with 100, 200, and 400 mg/kg of curcumin 10 days before and 2 days after $K_2Cr_2O_7$ injection, 2) pretreatment with 400 mg/kg of curcumin for 10 days before $K_2Cr_2O_7$ injection and 3) posttreatment with 400 mg/kg of curcumin 2 days after $K_2Cr_2O_7$ injection. Then Rats were sacrificed 48 hour after $K_2Cr_2O_7$ or vehicle injection to evaluate mitochondrial and renal function and oxidant stress. At the end of this study, complete treatment and pretreatment with curcumin ameliorated histological damage, renal dysfunction, oxidant stress and decrease in antioxidant enzyme activity in mitochondria and kidney, which were induced by $K_2Cr_2O_7$. Likewise, pretreatment with curcumin attenuated the mitochondrial dysfunction including changes in ATP content, oxygen consumption, mitochondrial membrane potential and calcium retention. Furthermore, curcumin pretreatment induced the nuclear translocation of the nuclear factor-E2-related factor 2 (Nrf2) and increased the activity of glutathione S-transferase, superoxide dismutase and glutathione reductase (67).

Sodium fluoride is an inorganic chemical compound with the formula NaF that can induce nephrotoxicity. Curcumin at the doses of 10 and 20 mg/kg body weight (intraperitoneally) revealed nephroprotective effects in a model of rats study. In this study, the pretreatment with curcumin one week before the administration of fluoride (600 ppm) was effective in normalization of serum urea, serum creatinine and blood urea nitrogen. Also, curcumin prevented the decreasing of antioxidant enzyme including catalase and superoxide dismutase and increasing in level of renal lipid peroxidation (68).

Conclusion

According to various epidemiological data, renal failure, either acute or chronic, is a serious health and economical problem throughout the world. The necessity to develop kidney protective modalities sets the eyes in compounds such as curcumin, which has been used in the traditional medicine, specifically because of its protective effects against kidney damage. In addition, curcumin is a free

radical scavenger due to its antioxidant capacity and many experimental investigations reported the ameliorative effect of curcumin in acute and chronic kidney problems such as diabetic kidney disease, nephrotoxicity and renal ischemia.

Authors' contribution

HN and ZAG wrote the primary draft. MRK edited the final manuscript.

Conflicts of interest

The authors declare no competing interests.

Ethical considerations

Ethical issues (including plagiarism, data fabrication, double publication) have been completely observed by authors.

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References

1. Ghosh SS, Gehr TW, Ghosh S. Curcumin and chronic kidney disease (CKD): major mode of action through stimulating endogenous intestinal alkaline phosphatase. *Molecules*. 2014; 19:20139-56.
2. Amini FG, Rafieian-Kopaei M, Nematbakhsh M, Baradaran A, Nasri H. Ameliorative effects of metformin on renal histologic and biochemical alterations of gentamicin-induced renal toxicity in Wistar rats. *J Res Med Sci*. 2012;17:621-5.
3. Rafieian-Kopaei M, Nasri H. R: recombinant human erythropoietin reduces rhabdomyolysis-induced acute renal failure in rats. *Injury*. 2013;44:1662.
4. Nasri H. World kidney day 2013: acute kidney injury; a public health aware. *Iran J Public Health*. 2013; 42:338-40.
5. Pickering JW, Endre ZH. The definition and detection of acute kidney injury. *J Renal Inj Prev*. 2013;3:21-5.
6. Guerrot D, Dussaule JC, Kavvadas P, Boffa JJ, Chadjiachristos CE, Chatziantoniou C. Progression of renal fibrosis: the underestimated role of endothelial alterations. *Fibrogenesis Tissue Repair*. 2012;5:S15.
7. Asouri M, Ataee R, Ahmadi AA, Amini A, Rezaei Moshaei M. Antioxidant and free radical scavenging activities of Curcumin. *Asian J Chem*. 2013;25:7593-7595.
8. Baradaran A, Nasri H, Rafieian-Kopaei M. Comment on: anti-oxidative stress activity of *Stachys lavandulifolia* aqueous extract in humans. *Cell J*. 2013;15:272-3.
9. Lahera V, Goicoechea M, de Vinuesa SG, Oubiña P, Cachofeiro V, Gómez-Campderá F, et al. Oxidative stress in uremia: the role of anemia correction. *J Am Soc Nephrol*. 2006;17:S174-7.
10. Barzegar A, Moosavi-Movahedi AA. Intracellular ROS protection efficiency and free radical-scavenging activity of curcumin. *PLoS One*. 2011;6:e26012.
11. Galle J. Oxidative stress in chronic renal failure. *Nephrol Dial Transplant*. 2001;16:2135-7.
12. Rafieian-Kopaei M, Nasri H. R: Erythropoietin ameliorates oxidative stress and tissue injury following renal ischemia/reperfusion in rat kidney and lung. *Med Princ Pract*. 2014;23:95.
13. Hammad FT, Al-Salam S, Lubbad L. Curcumin provides incomplete protection of the kidney in ischemia reperfusion

- injury. *Physiol Res*. 2012; 61(5):503-11.
14. Ardalan MR, Nasri H, Rafieian-Kopaei M. Comment on: Protective role of recombinant human erythropoietin in kidney and lung injury following renal bilateral ischemia-reperfusion in rat model. *Int J Prev Med*. 2013;4:1226-7.
 15. Ozbek E. Induction of oxidative stress in kidney. *Int J Nephrol*. 2012;2012:465897.
 16. Lobo V, Patil A, Phatak A, Chandra N. Free radicals, antioxidants and functional foods: Impact on human health. *Pharmacogn Rev*. 2010;4:118-26.
 17. Sen S, Chakraborty R, Sridhar S, Reddy YSR, De B. Free radicals, antioxidants, diseases and phytomedicines: current status and future prospect. *Int J Pharm Sci Rev Res*. 2010; 3:91-100.
 18. Nasri H, Tavakoli M, Ahmadi A, Baradaran A, Nematbakhsh M, Rafieian-Kopaei M. Ameliorative effect of melatonin against contrast media induced renal tubular cell injury. *Pak J Med Sci*. 2014;30:261-5.
 19. Bhullar KS, Jha A, Youssef D, Rupasinghe HP. Curcumin and its carbocyclic analogs: structure-activity in relation to antioxidant and selected biological properties. *Molecules*. 2013;18:5389-404.
 20. Nasri H, Rafieian-Kopaei M. Kidney tubular cell protection; recent findings. *Iran J Pediatr*. 2014;24:781-9.
 21. Abedigheshlaghi Z, Monajemi R, Yahyaabadi S. Cytotoxic effect of aqueous and alcoholic extracts of *Pterocarya fraxinifolia* leaves on K562 cell line. *Zahedan J Res Med Sci*. 2014;16:1-5.
 22. Nasri H. Antioxidants for prevention of gentamicin-induced nephrotoxicity. *Iran J Kidney Dis*. 2014;8:1-2.
 23. Baradaran A, Rabiei Z, Rafieian M, Shirzad H. A review study on medicinal plants affecting amnesia through cholinergic system. *J HerbMed Pharmacol*. 2012;1:3-9.
 24. Nasri H, Madihi Y, Marikhi A. Commentary on: Effects of Cinnamon Consumption on Glycemic Status, Lipid Profile and Body Composition in Type 2 Diabetic Patients. *Int J Prev Med*. 2013;4:618-9.
 25. Nasri H, Rafieian-Kopaei M. Tubular kidney protection by antioxidants. *Iran J Public Health*. 2013;42:1194-1196.
 26. Radak Z, Zhao Z, Koltai E, Ohno H, Atalay M. Oxygen consumption and usage during physical exercise: the balance between oxidative stress and ROS-dependent adaptive signaling. *Antioxid Redox Signal*. 2013;18:1208-46.
 27. Baradaran A, Nasri H, Rafieian-Kopaei M. Oxidative stress and hypertension: Possibility of hypertension therapy with antioxidants. *J Res Med Sci*. 2014;19:358-67.
 28. Tian N, Thrasher KD, Gundy PD, Hughson MD, Manning RD Jr. Antioxidant treatment prevents renal damage and dysfunction and reduces arterial pressure in salt-sensitive hypertension. *Hypertension*. 2005;45:934-9.
 29. Namjoo A, Nasri H, Talebi-Juneghani A, Baradaran A, Rafieian-Kopaei M. Safety profile of *Carthamus tinctorius* L. in lactation: brain, renal and hepatotoxicity. *Pak J Med Sci*. 2013;29:378-383.
 30. Trujillo J, Chirino YI, Molina-Jijón E, Andérica-Romero AC, Tapia E, Pedraza-Chaverrí J. Renoprotective effect of the antioxidant curcumin: Recent findings. *Redox Biol*. 2013;1:448-56.
 31. Nasri H, Rafieian-Kopaei M. Protective effects of herbal antioxidants on diabetic kidney disease. *J Res Med Sci*. 2014; 19:82-3.
 32. Nasri H, Shirzad H. Toxicity and safety of medicinal plants. *J HerbMed Pharmacol*. 2013;2:21-22.
 33. Nasri H, Rafieian-Kopaei M. Medicinal plants and antioxidants: Why they are not always beneficial?. *Iran J Public Health*. 2014;43:255-257.
 34. Rafieian-Kopaei M, Baradaran A, Rafieian M. Plants antioxidants: From laboratory to clinic. *J Nephropathol*. 2013; 2:152-3.
 35. Nasri H. Effect of garlic extract on blood glucose level and lipid profile in normal and alloxan diabetic rabbits. *Adv Clin Exp Med*. 2013;22:449-50.
 36. Sewell RD, Rafieian-Kopaei M. The history and ups and downs of herbal medicine usage. *J HerbMed Pharmacol*. 2014;3:1-3.
 37. Nasri H, Ardalan MR, Rafieian-Kopaei M. Mechanistic impacts of medicinal plants in diabetic kidney disease. *Iran J Public Health*. 2014;43:1311-3.
 38. Rafieian-Kopaei M. Medicinal plants and the human needs. *J HerbMed Pharmacol*. 2012;1:1-2.
 39. Nasri H, Rafieian-Kopaei M. Medicinal plants and new concerns in statin consumption. *Iran J Public Health*. 2014; 42:1071-2.
 40. Rafieian-Kopaei M, Hosseini M, Shirzad H. Comment on: effect of pomegranate flower extract on cisplatin-induced nephrotoxicity in rats. *J Nephropathol*. 2014;3:121-3.
 41. Tuorkey MJ. Curcumin a potent cancer preventive agent: Mechanisms of cancer cell killing. *Interv Med Appl Sci*. 2014; 6:139-46.
 42. Bandyopadhyay D. Farmer to pharmacist: curcumin as an anti-invasive and antimetastatic agent for the treatment of cancer. *Front Chem*. 2014;2:113.
 43. Shetty D, Kim YJ, Shim H, Snyder JP. Eliminating the heart from the curcumin molecule: monocarbonyl curcumin mimics (MACs). *Molecules*. 2014;20:249-92.
 44. Ak T, Gülçin I. Antioxidant and radical scavenging properties of curcumin. *Chem Biol Interact*. 2008;174:27-37.
 45. Yarru LP, Settivari RS, Gowda NK, Antoniou E, Ledoux DR, Rottinghaus GE. Effects of turmeric (*Curcuma longa*) on the expression of hepatic genes associated with biotransformation, antioxidant, and immune systems in broiler chicks fed aflatoxin. *Poult Sci*. 2009;88:2620-7.
 46. Ye SF, Hou ZQ, Zhong LM, Zhang QQ. Effect of curcumin on the induction of glutathione S-transferases and NAD(P)H:quinone oxidoreductase and its possible mechanism of action. *Yao Xue Xue Bao*. 2007;42:376-80.
 47. Rushworth SA, Ogborne RM, Charalambos CA, O'Connell MA. Role of protein kinase C delta in curcumin-induced antioxidant response element-mediated gene expression in human monocytes. *Biochem Biophys Res Commun*. 2006; 341:1007-16.
 48. Chen B, Zhang DP, Gao W. Effect of curcumin on the expression of collagen type I protein and transforming growth factor-beta1 mRNA in pulmonary fibrosis rats. *Zhonghua Lao Dong Wei Sheng Zhi Ye Bing Za Zhi*. 2008; 26:257-61.
 49. Rafieian-Kopaei M, Behradmanesh S, Kheiri S, Nasri H. Association of serum uric acid with level of blood pressure in type 2 diabetic patients. *Iran J Kidney Dis*. 2014;8:152-4.
 50. Soetikno V, Suzuki K, Veeraveedu PT, Arumugam S, Lakshmanan AP, Sone H, et al. Molecular understanding of curcumin in diabetic nephropathy. *Drug Discov Today*. 2013;18:756-63.
 51. Nasri H. Re: effect of silymarin on streptozotocin-nicotinamide-induced type 2 diabetic nephropathy in rats. *Iran J Kidney Dis*. 2013;7:414-5.
 52. Nasri H, Rafieian-Kopaei M. Effect of vitamin D on insulin resistance and nephropathy in type 2 diabetes. *J Res Med Sci*.

- 2014;19:581-2.
53. Behradmanesh S, Horestani MK, Baradaran A, Nasri H. Association of serum uric acid with proteinuria in type 2 diabetic patients. *J Res Med Sci.* 2013;18:44-6.
 54. Baradaran A, Madihi Y, Merrikhi A, Rafieian-Kopaei M, Nasri H. Serum lipoprotein (a) in diabetic patients with various renal function not yet on dialysis. *Pak J Med Sci.* 2013;29:354-357.
 55. Nasri H. Re metformin revisited: a critical review of the benefit-risk balance in at-risk patients with type 2 diabetes. *Diabetes Metab.* 2013;39:375-6.
 56. Soetikno V, Watanabe K, Sari FR, Harima M, Thandavarayan RA, Veeraveedu PT, et al. Curcumin attenuates diabetic nephropathy by inhibiting PKC- α and PKC- β 1 activity in streptozotocin-induced type I diabetic rats. *Mol Nutr Food Res.* 2011;55:1655-65.
 57. Soetikno V, Sari FR, Sukumaran V, Lakshmanan AP, Harima M, Suzuki K, et al. Curcumin decreases renal triglyceride accumulation through AMPK-SREBP signaling pathway in streptozotocin-induced type 1 diabetic rats. *J Nutr Biochem.* 2013;24:796-802.
 58. Pan Y, Zhu G, Wang Y, Cai L, Cai Y, Hu J, et al. Attenuation of high-glucose-induced inflammatory response by a novel curcumin derivative B06 contributes to its protection from diabetic pathogenic changes in rat kidney and heart. *J Nutr Biochem.* 2013;24:146-55.
 59. Pan Y, Wang Y, Cai L, Cai Y, Hu J, Yu C, et al. Inhibition of high glucose-induced inflammatory response and macrophage infiltration by a novel curcumin derivative prevents renal injury in diabetic rats. *Br J Pharmacol.* 2012;166:1169-82.
 60. 60-Morigi M, Perico L, Rota C, Longaretti L, Conti S, Rottoli D, et al. Sirtuin 3-dependent mitochondrial dynamic improvements protect against acute kidney injury. *J Clin Invest.* 2015;125:715-26.
 61. Pabla N, Dong Z. Cisplatin nephrotoxicity: mechanisms and renoprotective strategies. *Kidney Int.* 2008;73:994-1007.
 62. Zhan M, Brooks C, Liu F, Sun L, Dong Z. Mitochondrial dynamics: regulatory mechanisms and emerging role in renal pathophysiology. *Kidney Int.* 2013; 83:568-81.
 63. Brooks C, Wei Q, Cho SG, Dong Z. Regulation of mitochondrial dynamics in acute kidney injury in cell culture and rodent models. *J Clin Invest.* 2009;119:1275-85.
 64. Bayrak O, Uz E, Bayrak R, Turgut F, Atmaca AF, Sahin S, et al. Curcumin protects against ischemia/reperfusion injury in rat kidneys. *World J Urol.* 2008;26:285-91.
 65. Tarasub N, Tarasub C, Devakul Na Ayutthaya W. Protective role of curcumin on cadmium-induced nephrotoxicity in rats. *J Environ Chem Ecotoxicol.* 2011;3:17-24.
 66. Agarwal R, Goel SK, Behari JR. Detoxification and antioxidant effects of curcumin in rats experimentally exposed to mercury. *J Appl Toxicol.* 2010;30:457-68.
 67. Molina-Jijón E, Tapia E, Zazueta C, El Hafidi M, Zatarain-Barrón ZL, Hernández-Pando R, et al. Curcumin prevents Cr(VI)-induced renal oxidant damage by a mitochondrial pathway. *Free Radic Biol Med.* 2011;51:1543-57.
 68. Nabavi SF, Moghaddam AH, Eslami S, Nabavi SM. Protective effects of curcumin against sodium fluoride-induced toxicity in rat kidneys. *Biol Trace Elem Res.* 2012;145:369-74.

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